PrimePrep™ Gel Purification Kit

[지금 받으신 제품은 Sample size로써, 10회 prep.을 하실 수 있습니다.
Buffer PW에 첨가할 Ethanol의 양은 bottle label에 표기되어있습니다.]

Introduction

*PrimePrep*TM Gel Purification Kit offer simple, rapid and cost-effective method for purification from agarose gel in TAE or TBE buffer system..
The purified DNA can be directly used in ligation, sequencing and other downstream application.

Kit Components

Reagents \ Cat. No.	K-8000 (50 prep.)
Spin column	50 ea
Buffer Gel-B	60 ml (30 ml x 2)
Buffer PW	10 ml
Buffer PE	10 ml

Before you begin

- **Add ethanol to Buffer PW before use.**

→ bottle의 label에 첨가량이 표시되어 있습니다.

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Experimental Protocol

1. **Cut out the interesting DNA fragment with a sharp scalpel or razor blade.**
   Minimize gel volume by cutting gel slice as small as possible.
2. **Weigh the gel slice in a microcentrifuge tube. Add 3 volumes of Buffer Gel-B to 1 volume of gel.**
   If the 100 mg of agarose gel slice, add 300 ul of Buffer Gel-B.
   If more than 1.5% agarose gel, add 6 volumes of Buffer Gel-B.
3. **Vortex the mixture and incubate at 55℃ until the agarose gel is completely melted (5~10min). To help the dissolving of gel, vortex the tube every 2~3 min during the incubation. Centrifuge the tube briefly at room temperature.**
4. **(Optional) Add 1 gel volume of isopropanol toe the sample and vortex to mix.**
   Do not centrifuge after mixing well.
   This step increases the yield of DNA fragment ＜200bp or ＞5kb.
5. **Transfer the mixture to a Spin column.**
6. **Centrifuge for 1 min at 13,000 rpm. Discard the flow-through and re-inserting the spin column to the collection tube.**
7. **(Optional) Add 500 ul of Buffer Gel-B to the column and centrifuge for 30 sec at 13,000 rpm. Discard the flow-through and re-inserting the spin column to the collection tube.**
   This step is complete removal of agarose for very sensitive applications.
8. **Add 700 ul Buffer PW and centrifuge for 30 sec. at 13,000 rpm.**
   Discard the flow-through and re-inserting the spin column to the collection tube.
9. **Centrifuge for an additional 1 min at 13,000 rpm to remove residual wash buffer.**
   Residual ethanol of washing buffer may inhibitor subsequent enzymatic reaction.
10. **Transfer the spin column to new 1.5 ml microcentrifuge tube.**
    The 1.5 ml microcentrifuge tube is not provided.
11. **Add 50 ul of Buffer PE or deionized distilled water to the center of the membrane in the column, let stand for 1 min and centrifuge for 1 min at 13,000 rpm.**
    For larger fragment(＞5kb), use pre-warmed (70℃) Buffer PE for best efficiency.

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